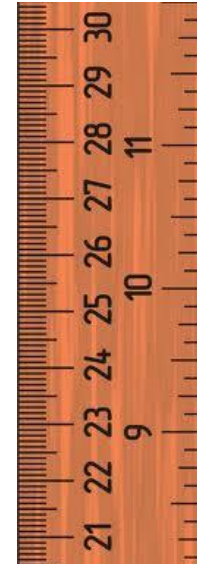
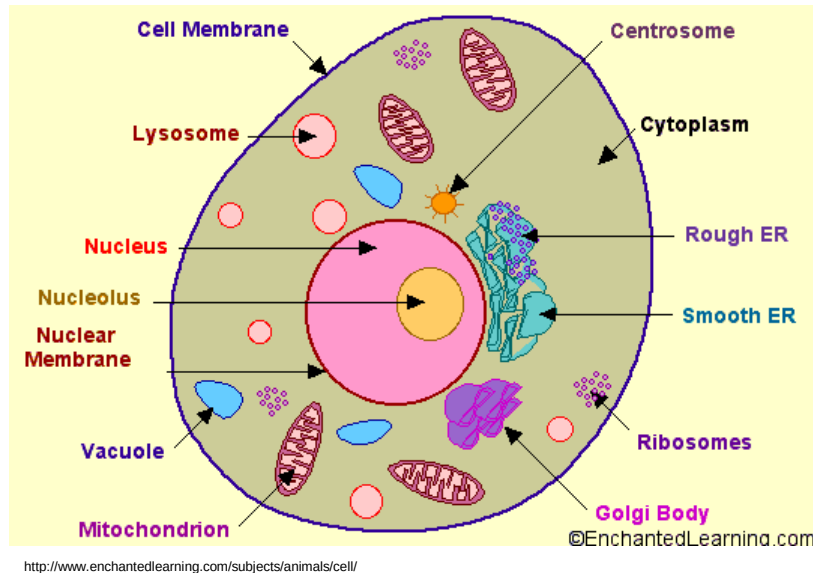


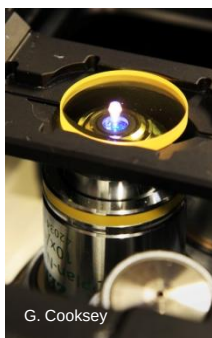
Issues in validating cell-imaging based assays



J. Elliott, A. Plant, M. Halter, G. Cooksey, K. Bhadriraju, A. Tona
Cell System Sciences Group
NIST Gaithersburg
AIMBE/NIH Summit, March 19, 2012

What is NIST....

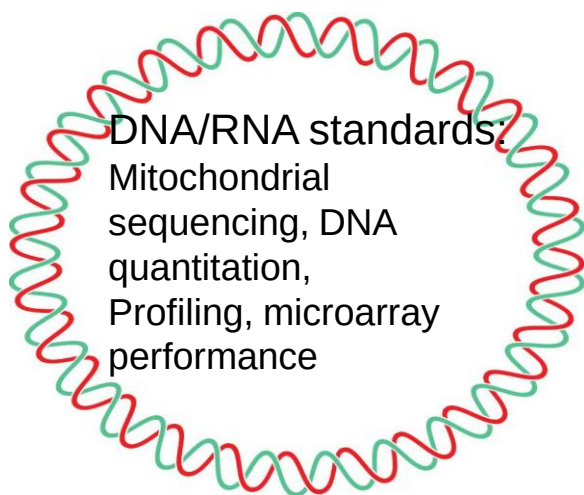
- Department of Commerce
- Mission to improve American life via good measurements
- Standards, new instrumentation and new measurement strategies



Optical standards:
ND Filter, fluorescence,
spectral



Chemical standards:
Vitamin D, hormone in
serum, albumin



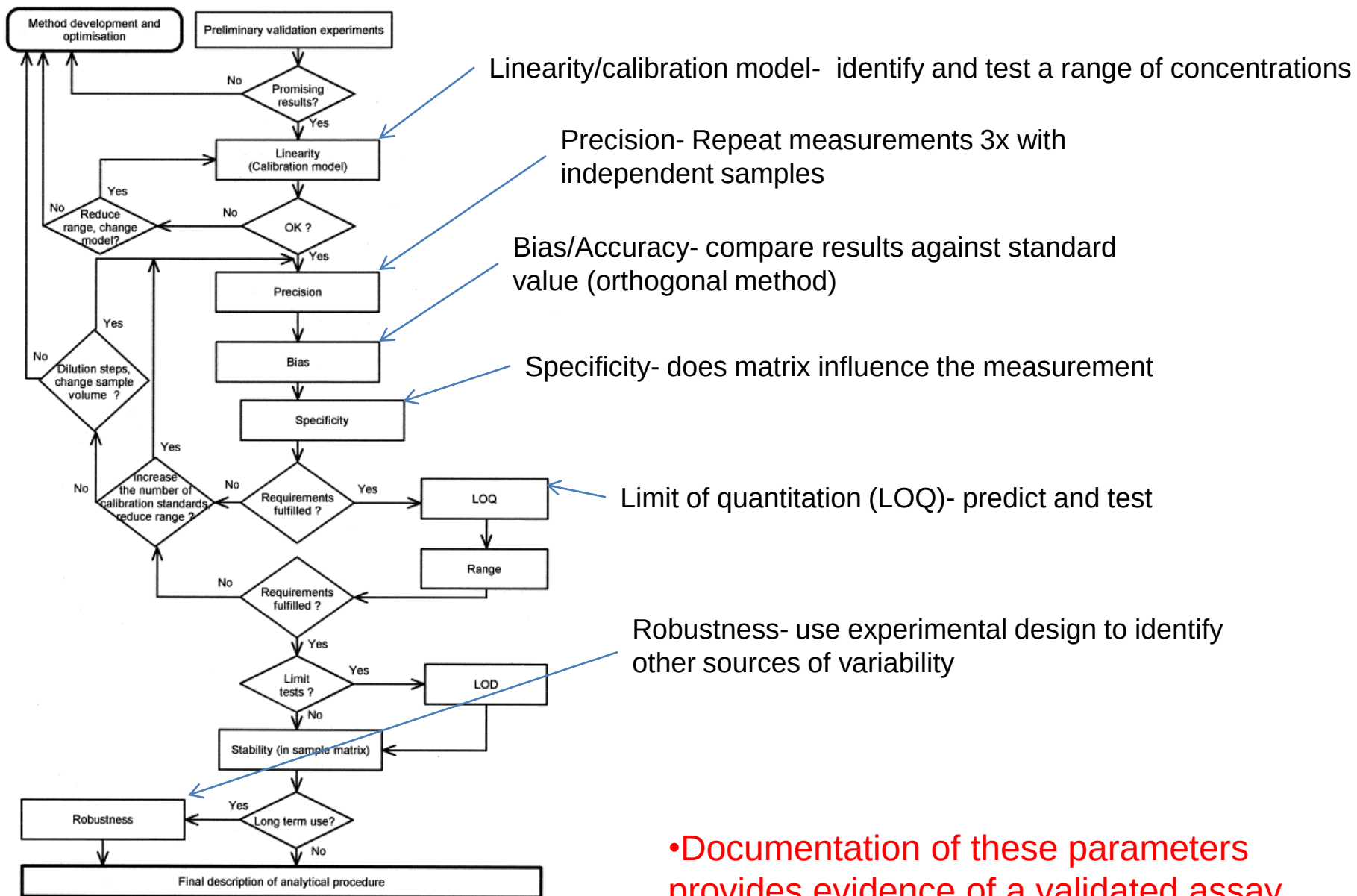
Documentary standards

Validation of a measurement

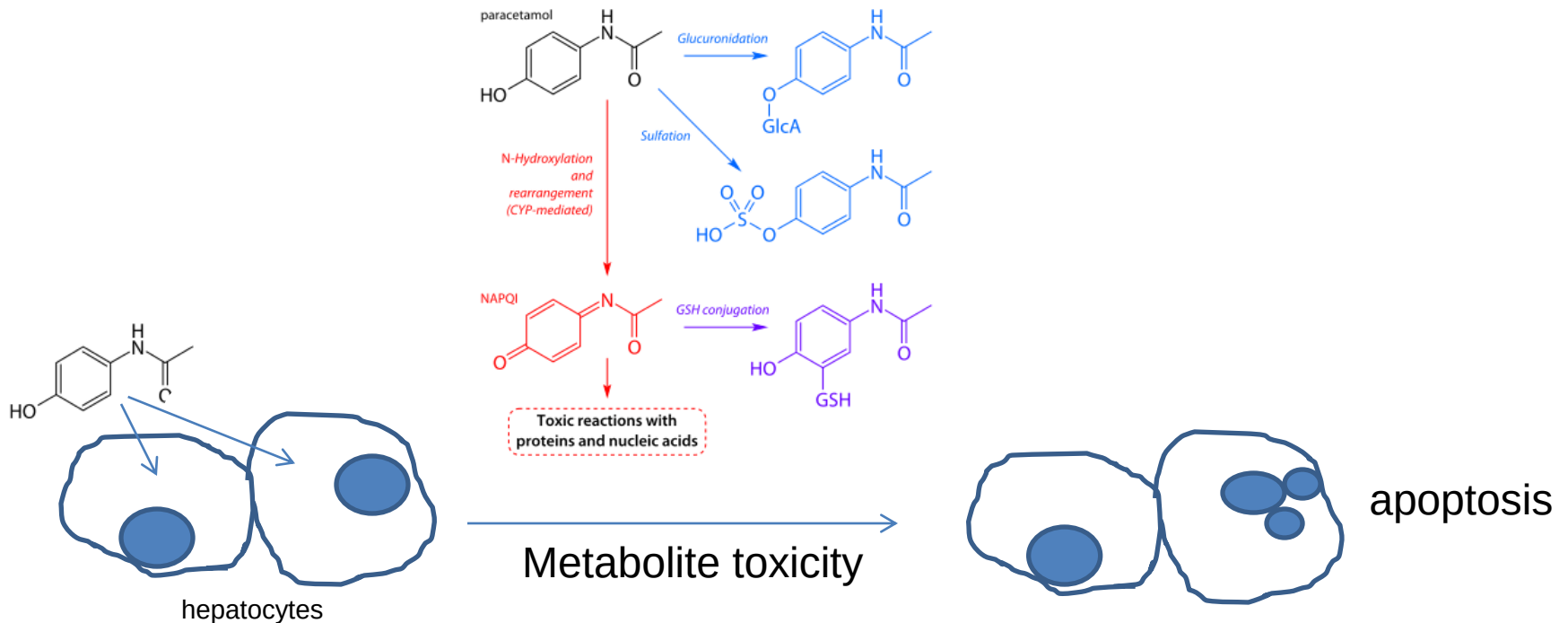
Is the measurement right? What's the evidence?

- Measurement characteristics
 - **Accuracy**- How close to the real value?
 - **Precision**- how much random error?
 - **Response function**- linearity/calibration
 - **Reproducibility**-do you get the same measurement next week?
 - **Robustness**-sensitivity to variations in assay parameters
- Instrument performance characteristics
 - **Dynamic range**- lowest limit of detection, highest limit of detection
 - **Contrast transfer function**- signal to noise, resolution
- Standards, reference materials and protocols
 - Reference materials- flatfield standard, fluorescence standard
 - Quantitative imaging protocol (i.e. flatfield, dark counts, etc)
 - Positive and negative controls, calibration curve

Example: Bioanalytical Validation flowchart (analyte concentration)



Validation of biological function

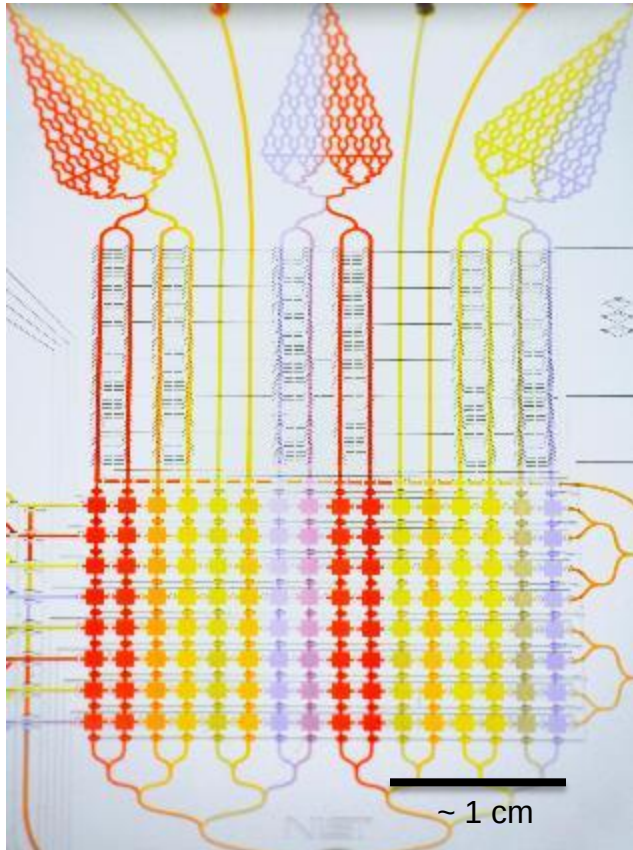


- Is the biological mechanism of action true?
 - Control compounds with known effects (inhibitors, activators)
 - Additional high quality assays and experiments likely required (live cell imaging, knockouts)
 - The aggregate of data validates a biological function.

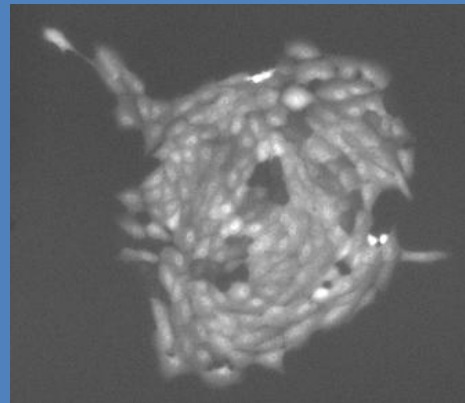
Microfluidic cellular assays

- Advantage: Complex plumbing possible, highly parallel, less reagent
- Disadvantage: not the same as “bulk” TCPS dish gold standard- high surface to volume ratio (leaching), gas permeability, cell seeding

128 chamber device w/ gradient mixer (G. Cooksey)



Ribosome Inhibitor Assay



CMV-dsEGFP-Vero cells
 $t_{1/2} \sim 2h$

Validation Issues

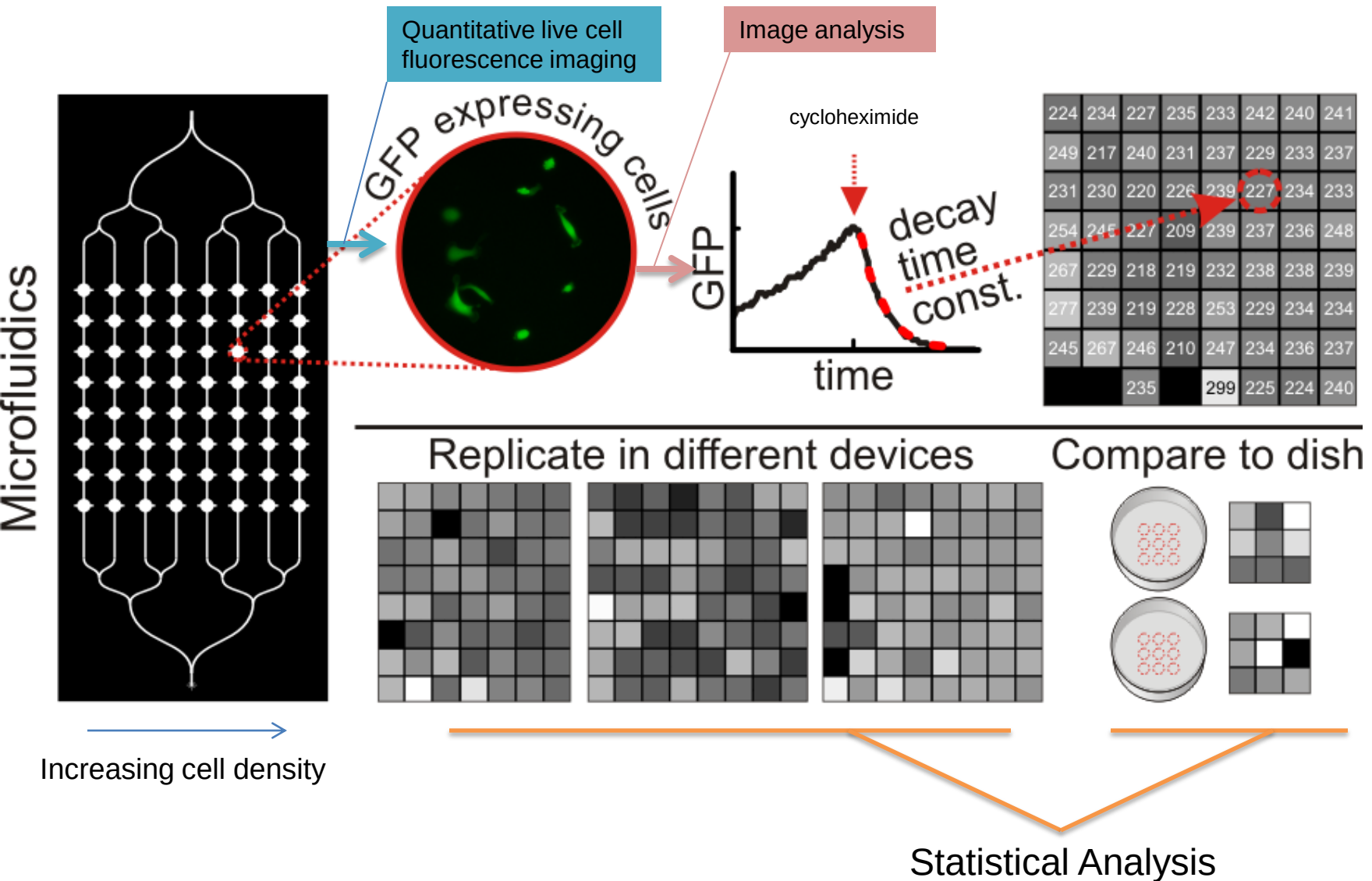
Results comparable to TCPS?

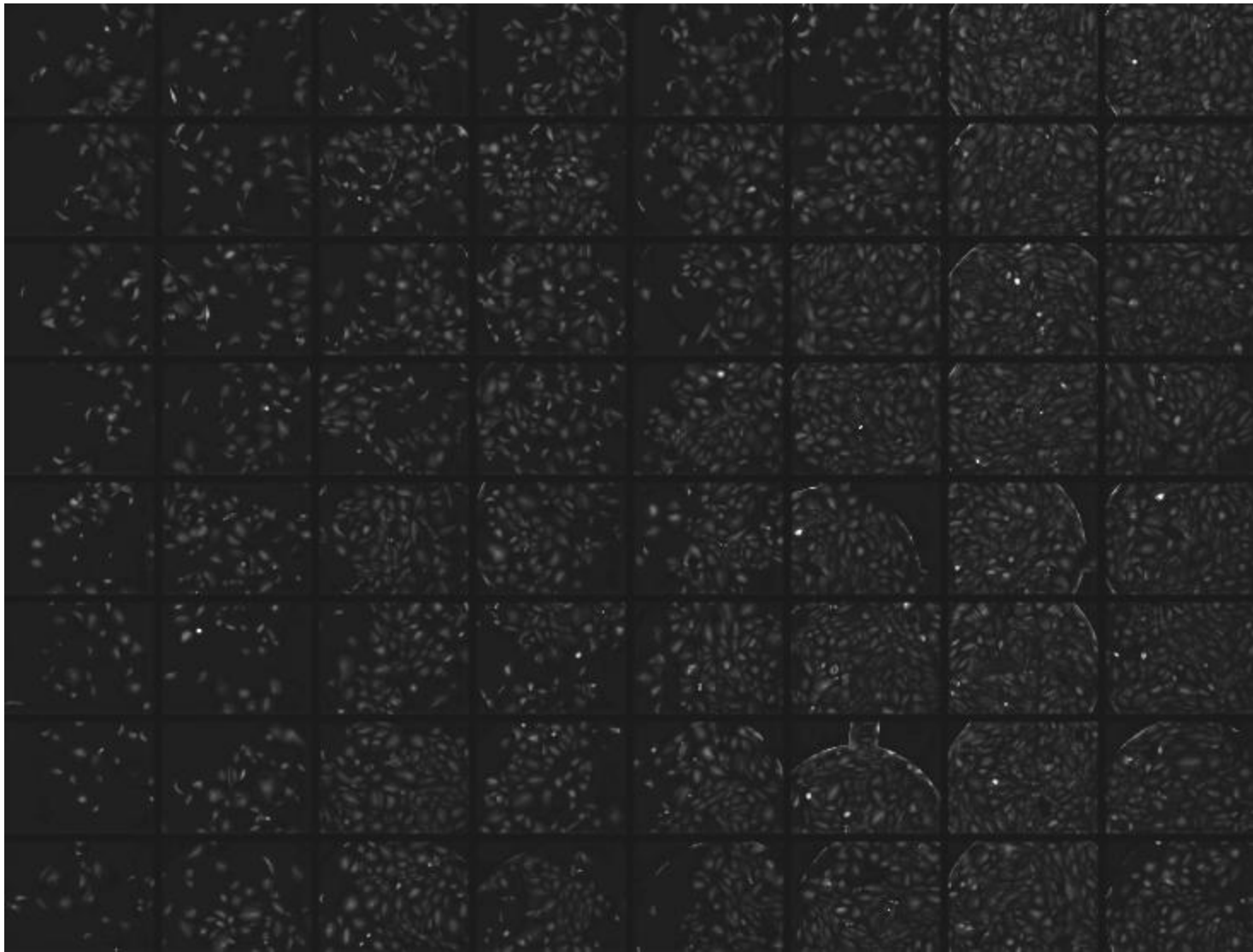
Cell density effects?

Paracrine signaling effects?

Flow rate, tubing fabrication,
imaging platform effects?

Experimental Design for Device Performance Evaluation





dsEGFP-Vero cells and cycloheximide- originally developed for ricin potency

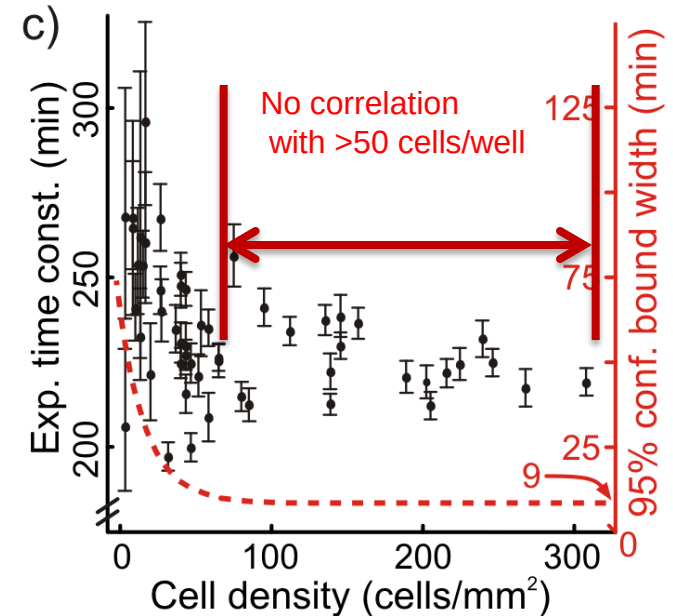
GFP Decay Assay Performance Summary

Substrate	Tubing	Lamp	Flow (ul/min)	Mean $T \pm \text{stdev}$ (min)	n (# regions)	Signal-to-Noise Ratio
Microfluidic Experiment A	Teflon	LED	0.1-1.5	236 ± 27	54	64 ± 2
Microfluidic Experiment B	Teflon	Hg arc	0.5-3.5	224 ± 31	64	47 ± 3
Microfluidic Experiment C	Tygon [®]	Hg arc	0.5-2	236 ± 15	61	48 ± 3
Microfluidic* Experiment D	Tygon [®]	Hg arc	0.5-2	254 ± 19	61	138 ± 50
TCPS dish	N/A	LED	Bulk	244 ± 66	8	14 ± 3
A		Hg arc		239 ± 48	9	7 ± 1
B		Hg arc		215 ± 27	9	9 ± 1
C		Hg arc		250 ± 13	9	9 ± 2
D	N/A	LED	Bulk	254 ± 41	9	11 ± 1
A		Hg arc		246 ± 13	9	11 ± 2
B						

N/A = not applicable

Bulk = 3ml stagnant media

*mean $\pm$ std (5 to 15h)



Questions:

Evidence

Comparable to TCPS?

Yes. No statistical difference.

Cell density effects?

No after 50 cells/well

Well position effects?

No. No statistical difference.

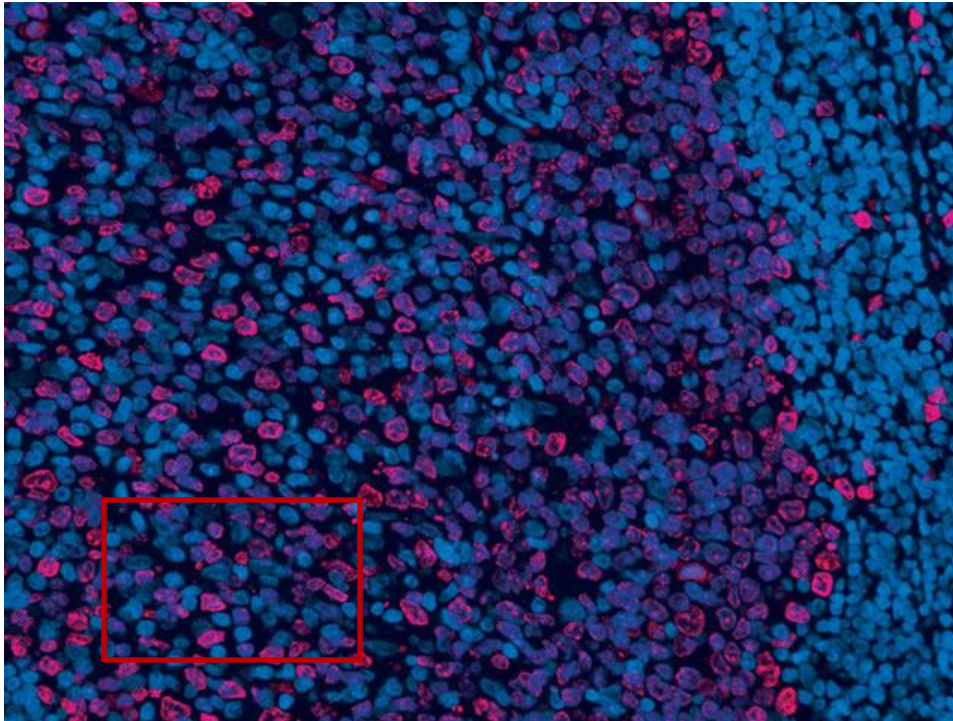
Robust?

Robust to fabrication, tubing, lamp, flow rate, replication

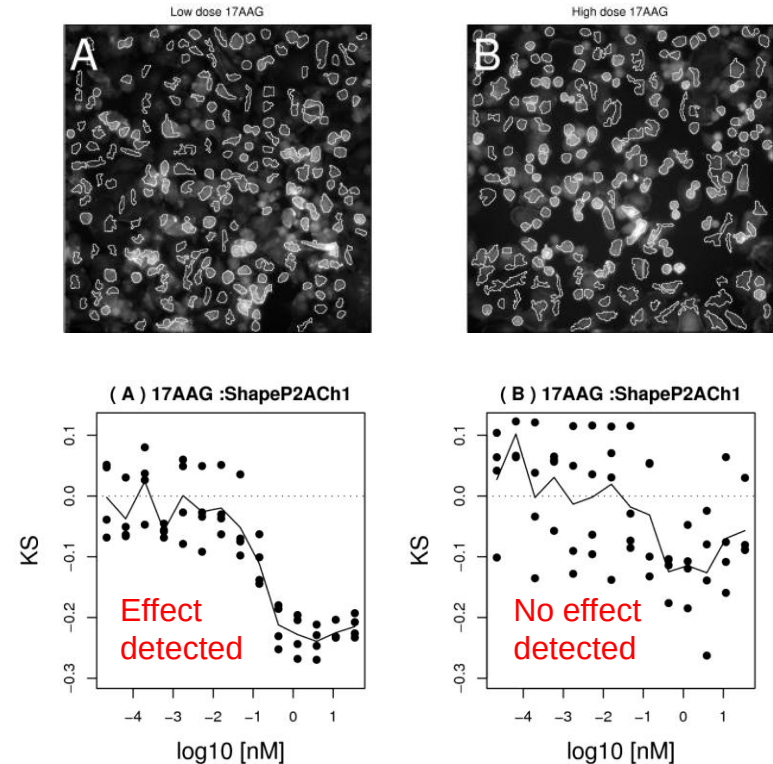
Other information:

5-fold higher signal to noise

Evaluating the Accuracy of Image Analysis Tools



Tonsil tissue w/DAPI and Ki-67 antibody (Biocare Medical)

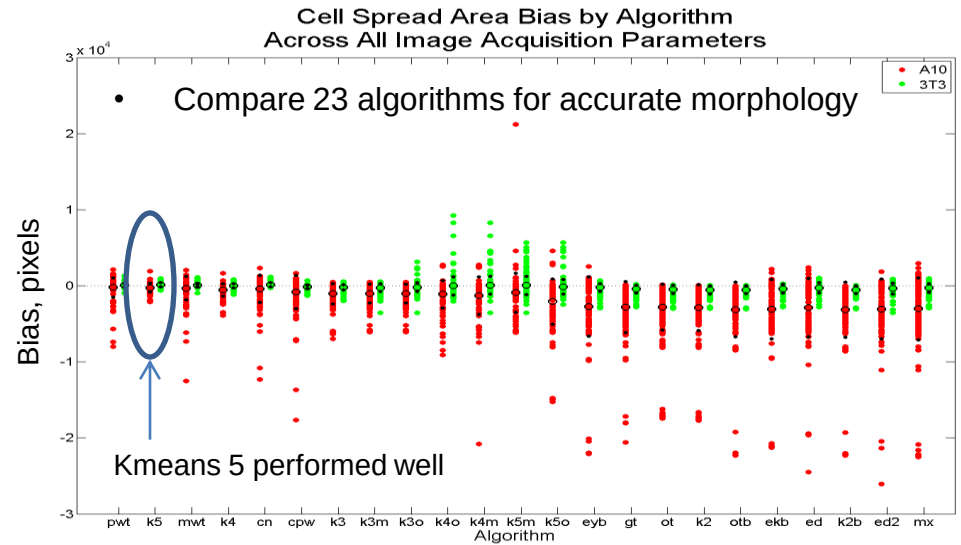
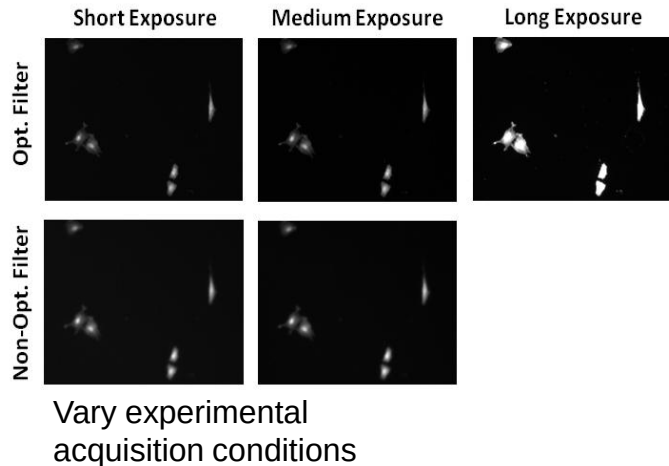


Hill et al BioMed Central Bioinformatics 2007

- What do you want to quantify (metric)?
 - How many images are required?
 - What image analysis do you use?
 - What validation evidence do you have?
- Inaccurate image analysis can lead to erroneous conclusion!!

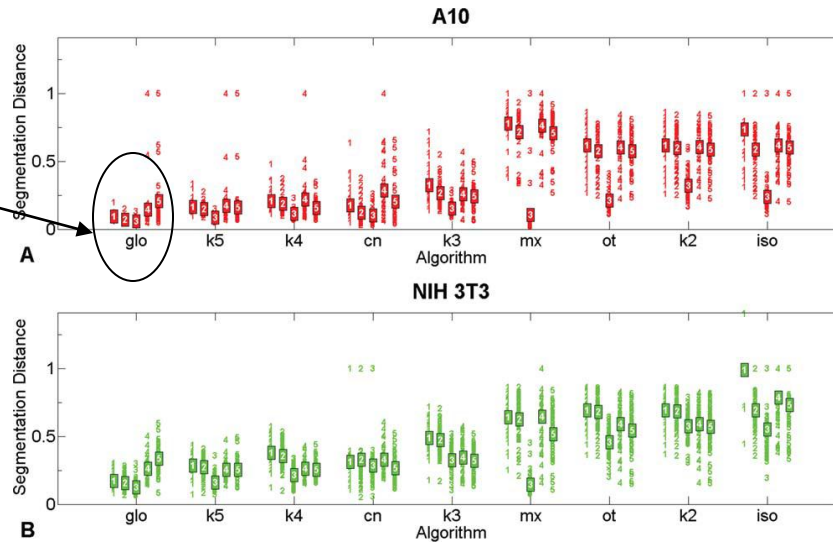
Evaluation the Accuracy of Cell Segmentation

Fluorescent images



Dima et al Cytometry A 2011

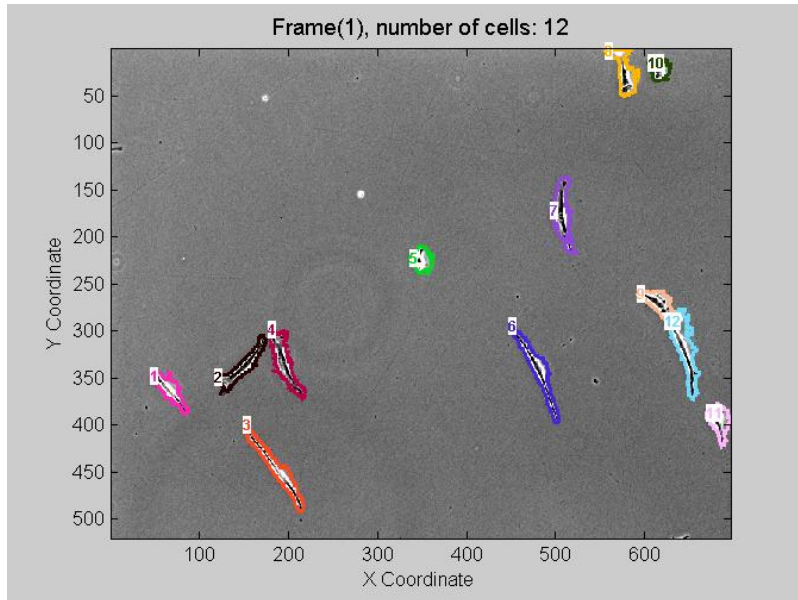
Robust over imaging conditions



- Requires manual (expert-trained) segmentation data to serve as reference data
- Image series, benchmarks, manual segmentation databased at www.sbd.gov

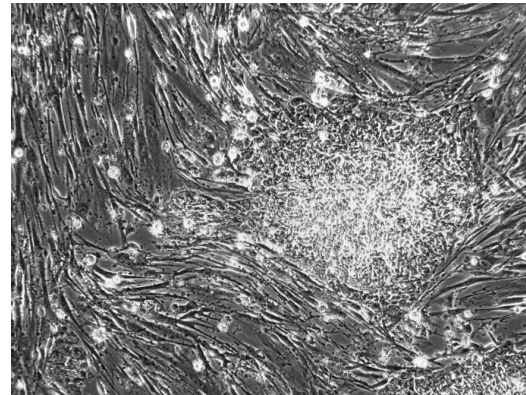
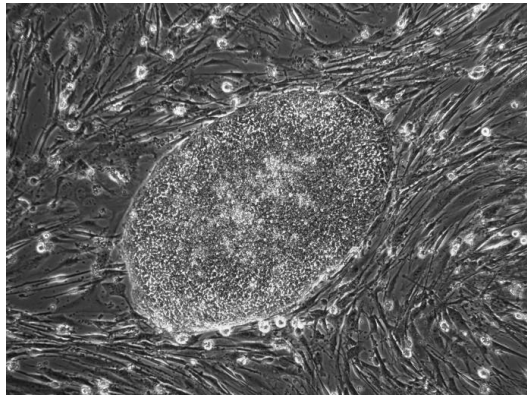
Non-destructive Phase Imaging

Phase imaging, NIH 3T3



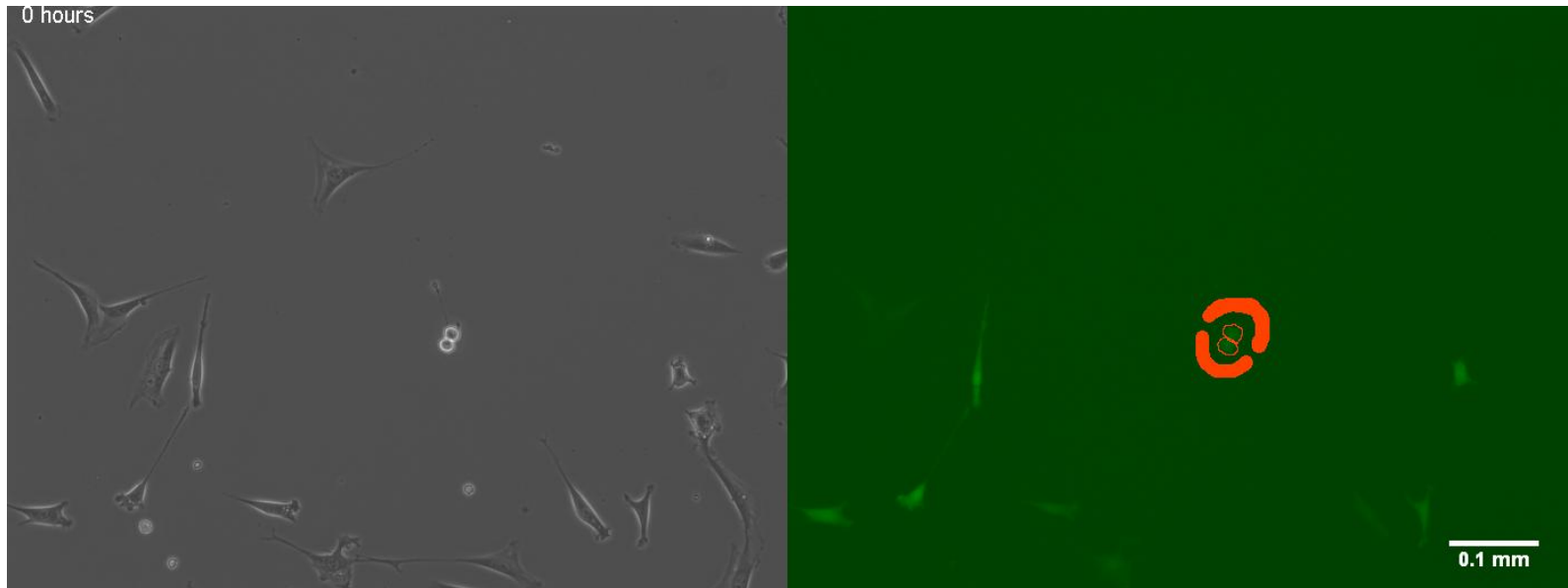
- Phase can be used for live cell imaging.
- Segmentation algorithm custom
- Need to determine acceptance criteria

Pluripotent
stem cells



Differentiation in
stem cells

Quantifying GFP reporter activity in live cells



CTRL: Flatfield corrected with 50% fluorescein solution

CTRL: <3% photobleaching of GFP over imaging time

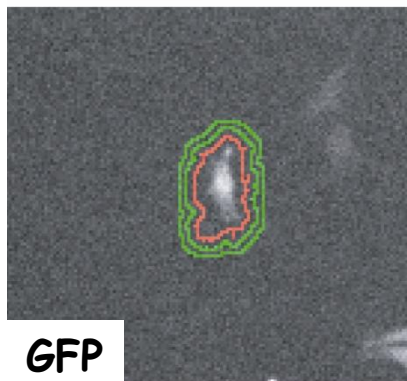
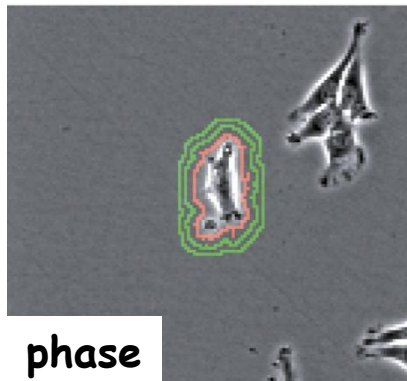
CTRL: Stable lamp intensity (LED)

97% complete-cell-cycle accuracy (193 tracked cells out of 199)

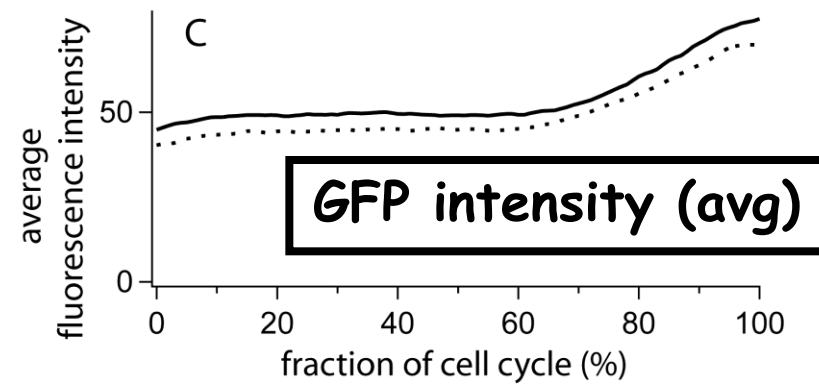
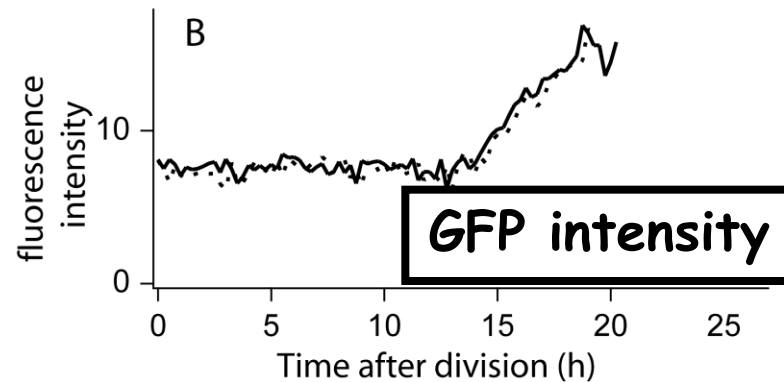
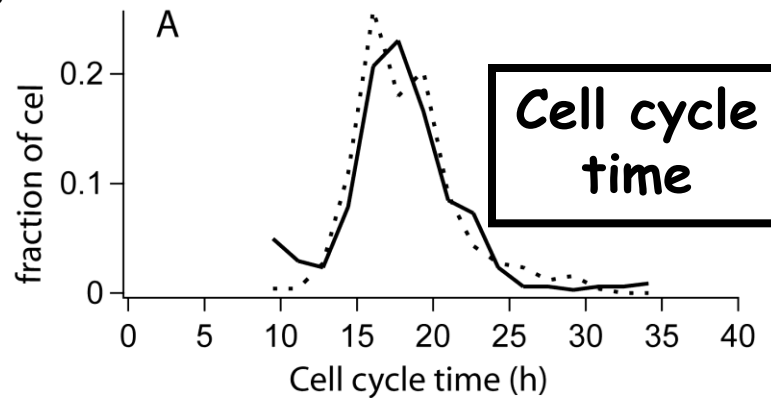
99.92% frame-to-frame accuracy (6 missed tracks out of 7957)

Validation of automated algorithms

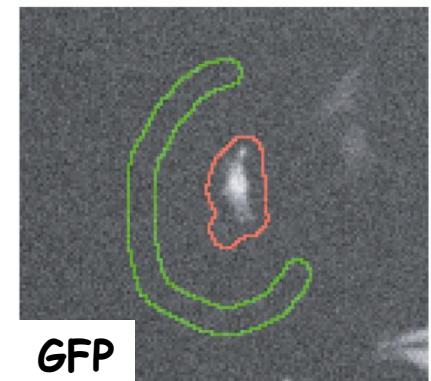
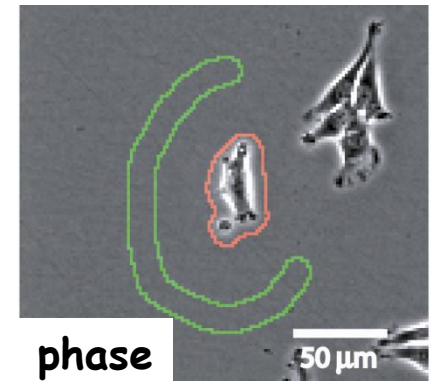
Automated analysis n=344



Adjusted Rand
Index
0.956
(7,186 masks)



Manual analysis n=257



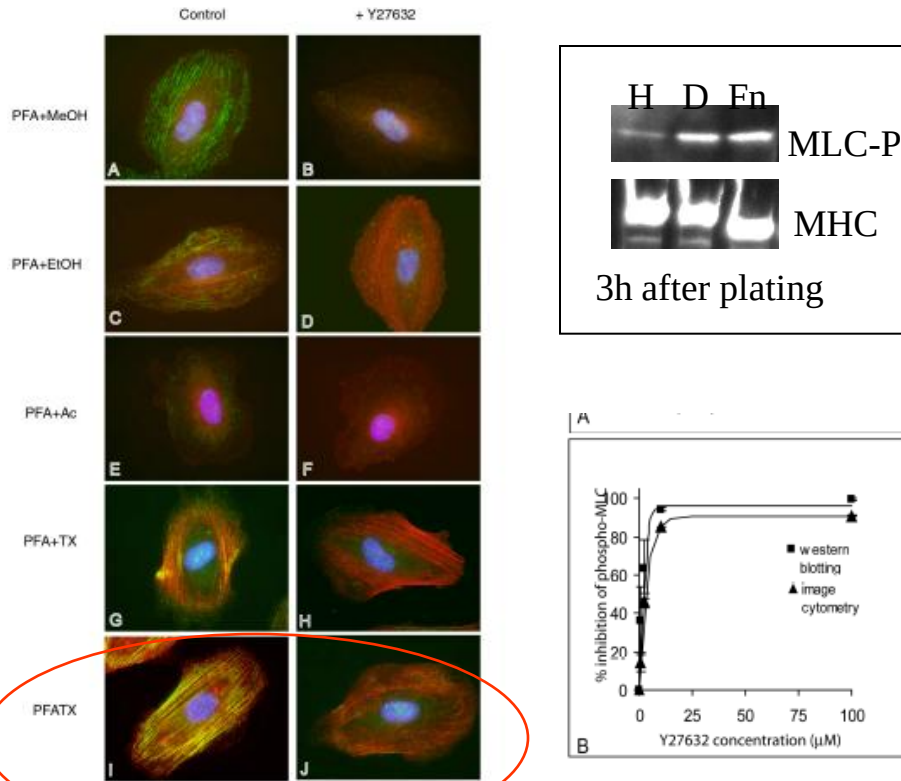
Cell cycle times: 17.1
h $\pm$ 4.3 h (man); 17.3
h $\pm$ 4.3 h

Halter et al. *Cytometry A*, 2011

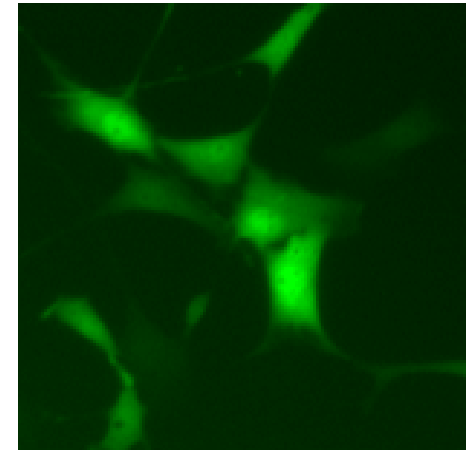
Validating Cell Imaging Protocols

Effects of Fixation

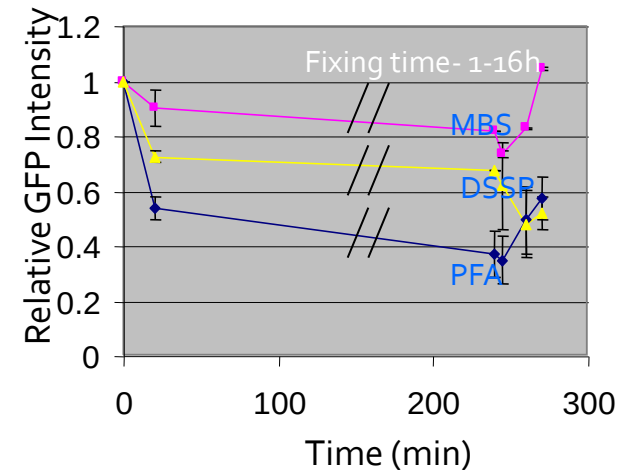
Phospho-myosin



Cytoplasmic GFP



In situ cell-by-cell analysis



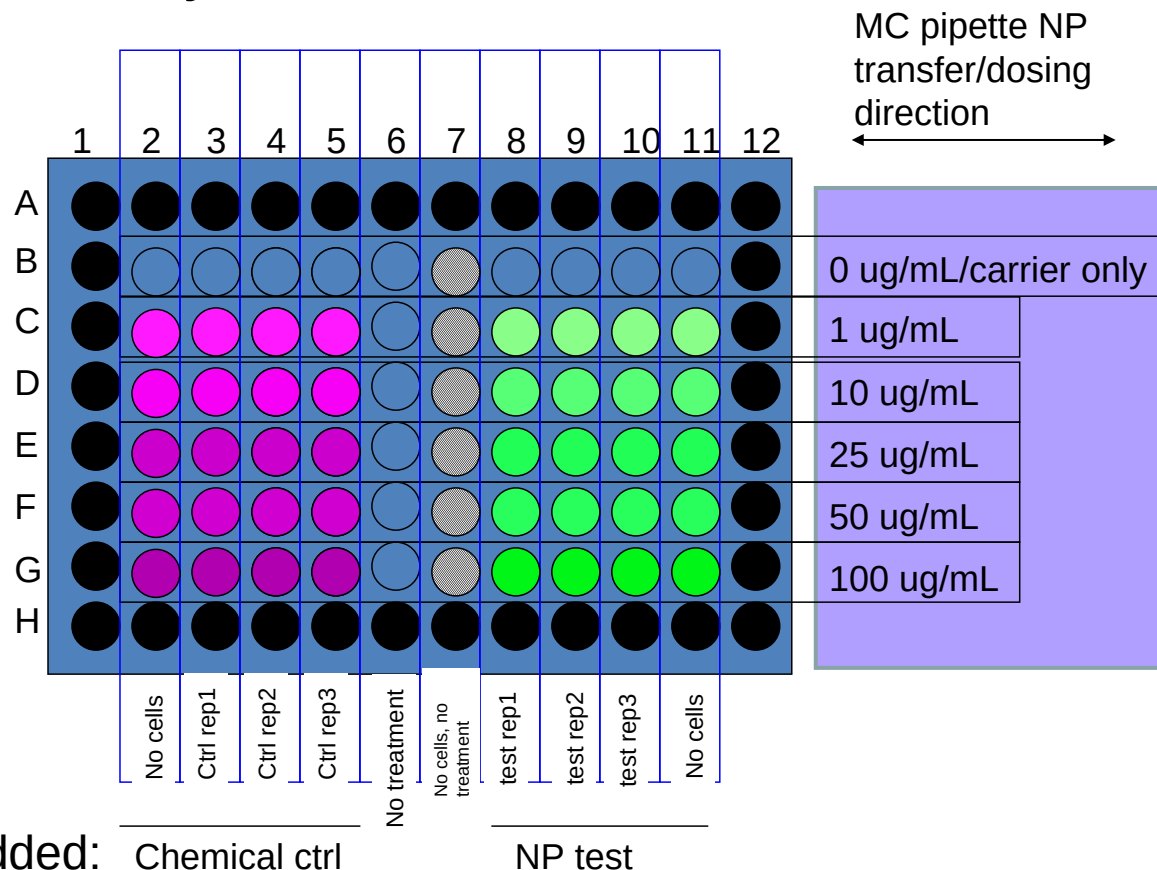
Redesigning Cell-based Assays with Validation Criteria: Nanocytotox



International Alliance for
NanoEHS Harmonization

- Stage 3.1 (2010)- Lack of agreement in interlaboratory testing (9 expert academic labs)

•Why??



- Several control experiments added:

- within pipette variability
- between pipette step variability
- collection of raw absorption data
- no cell NP test

- Protocol modified:

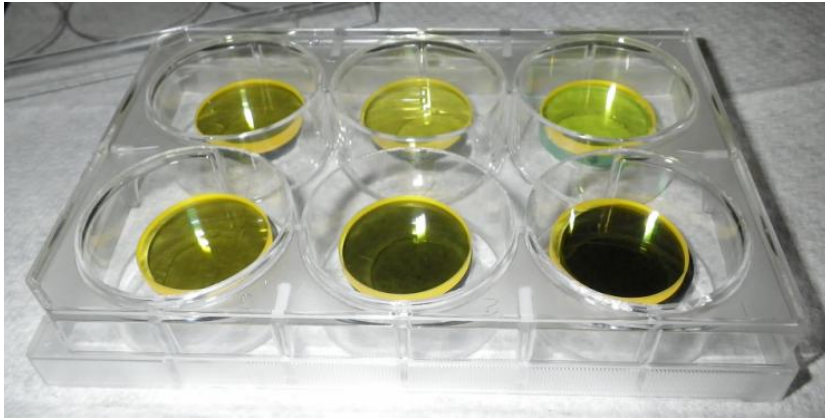
- directed pipette procedure
- directed dilution procedure
- directed NP dispersion procedure

Validation criteria:

- within pipette variability <6%
- between pipette step variability <10%
- OD <1.2 in no treatment wells
- reagent only well variability <3%
- EC50 CdCl₂ ctrl ~30 uM
- no absorption in NP only wells

Instrumentation Benchmarking Tools

Fluorescence Imaging System

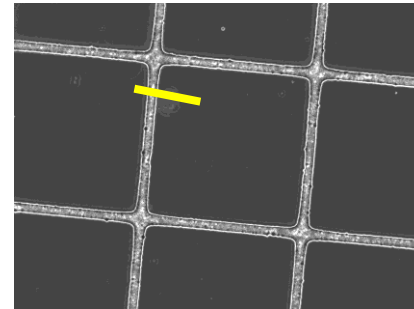


Prototype fluorescent microscope benchmarking tool- Halter 2012

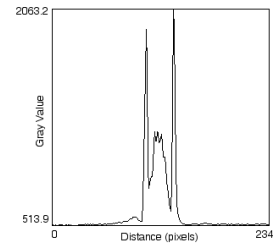
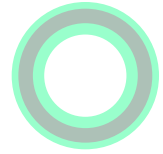
- Photostable fluorescent glass
- Dynamic range
- Signal to noise
- Lower limit of detection
- Upper limit of detection (saturation)

Phase Microscopy

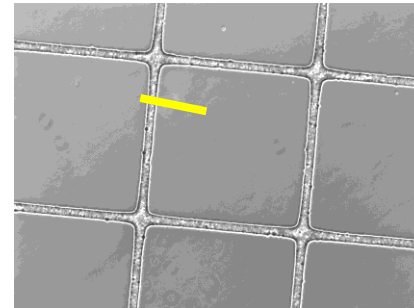
Correct alignment



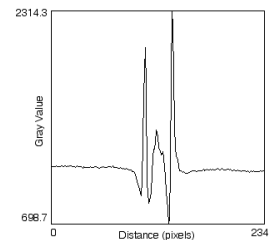
PDMS stamp on glass



Incorrect alignment



Halter et al 2011



- Need for benchmarking tools and protocols to specify the quality of an imaging system

Summary

- Need quantitative measurements to assess the quality of an assay method
 - Accuracy, precision, robustness, reproducibility
- Requires reference data, reference materials (standards), benchmarks, controls, statistics
- Experimental design can be used to establish robustness to factors
 - Fabrication, culture conditions, imaging system, etc.
- Cell imaging requires validation of image analysis, cell preparation protocols and benchmarking of imaging conditions
- Specifications that must be met before an assay is deemed valid can be identified with experimental design and quantitative measurements.